

Journal of Chromatography A, 814 (1998) 253-260

JOURNAL OF CHROMATOGRAPHY A

Short communication

Group identification of essential oils components using partition coefficients in a hexane-acetonitrile system

V.A. Isidorov^a, I.G. Zenkevich^b, E.N. Dubis^{a,*}, A. Slowikowski^a, E. Wojciuk^a

^aInstitute of Chemistry, Warsaw University, Bialystok Branch, Al. Pilsudskiego 11/4, 15-443 Bialystok, Poland ^bInstitute of Chemistry, St. Petersburg University, 198904 St. Petersburg, Russia

Received 17 December 1997; received in revised form 4 May 1998; accepted 5 May 1998

Abstract

Partition coefficients K_p in a hexane-acetonitrile heterogeneous system for 70 components usually encountered in essential oils of plants are reported. It is shown that the application of these values in the group identification stage greatly increases its reliability. The database for K_p values can be extended by calculations on the basis of the solubility-structure or retention-structure relationships. © 1998 Elsevier Science B.V. All rights reserved.

Keywords: Oils; Partition coefficients; Retention indices; Retention-structure relationships; Terpenes

1. Introduction

The problem of identification reliability has not yet lost its importance, although informative methods for the analysis of complex mixtures of organic compounds such as capillary gas chromatography in combination with mass spectral or Fourier transform infrared detector (FTIR) have been developing very rapidly. This problem is especially topical for the investigation of mixtures consisting of numerous components belonging to different classes of organic compounds. In particular, considerable difficulties are encountered in the so-called 'prospective' analyses of air, water or soil pollutants in which the a priori information on sample composition often does not exist [1]. The same can be said of analysis of essential oils.

Gas chromatographic (GC) identification is mainly

based on using retention indices (I). In recent years a considerable improvement in the reproducibility of characteristics of GC columns with different polarities has been observed. Therefore, the reproducibility of retention parameters reported by different authors has also improved. However, I values are one-dimensional analytical parameters, and their use often leads to ambiguous identification results. Identification precision can be increased by applying additional characteristics, such as the difference in I values for columns with different polarity [2]. The coefficients of partition of the analytes between two immiscible liquids, $K_p = C_1/C_2$, have also long been recommended as such parameters. The approach based on extra column phase equilibrium (its theory has been considered in a monograph [3]) has been used successfully for the analysis of pesticides [4-7]. Nevertheless, it has not been widely used so far. This is caused to a considerable extent by the absence of representative databases for K_{p} values. However, it

^{*}Corresponding author.

^{0021-9673/98/\$19.00 © 1998} Elsevier Science B.V. All rights reserved. PII: S0021-9673(98)00398-7

has recently been shown that the combination of I and K_p values ensures the efficiency of group identification, which is actually not inferior to that attained by using the differences in I values on columns with different polarities [8].

This work reports on the results of K_p determinations for a large number of terpene compounds in a hexane-acetonitrile system in the analysis of essential oils for ten species of plants. It is shown that K_p values can be predicted on the basis of retention-structure relationships.

2. Experimental

2.1. Materials

Essential oils of ylang-ylang, lavender, bergamot, lemon, and tangerine of Misitano & Stracuzzi (Italy) were used. Essential oils of pine (Pinus sylvestris), spruce (Picea abies), larch (Larix europeae), juniper (Juniperus communis), and thuja (Thuja occidentalis) were obtained in the laboratory of the Institute of Chemistry of Warsaw University (Bialystok Branch) by hydrodistillation from plants growing in the Bialystok region. Moreover, to determine K_{p} values, the preparations of borneole, isoborneole, linaloole, α -terpineole, fenchone, citral (isomers mixture), and α -terpenyl acetate from the collection of the Institute of Chemistry of St. Petersburg University (Russia) were used. Samples of andrane (α -cedrene epoxide), piconia (isolongifolanone), bisabolone, bisabolol, α,β -santalols, farnesol, nerolidol and cedranoxyde were obtained from a factory of aromatic substances (Grodzisk Mazowiecki, Poland).

Acetonitrile (HPLC grade, Merck), *n*-hexane, and toluene were used without additional purification or drying.

2.2. Sample preparation and analysis

Samples were prepared for analysis at room temperature. A 3-ml flask was charged through a pipette with 0.5 ml of *n*-hexane and 0.5 ml of acetonitrile and 30 μ l of essential oil (or hexane solution of a terpenoid from the collection), 1 μ l of

toluene and 5 μ l of C₈-C₁₄ or C₈-C₁₈ *n*-alkanes were added. Subsequently the flask was shaken for 30 s, and after phase separation each phase was subjected to GC analysis. Components were separated with flow splitting (1:30) on a GC DANI 86.10 HT chromatograph (Instrumentazione Analitica, Monza, Italy) with a flame ionisation detection (FID) system on fused-silica columns (30 m×0.25 mm, film thickness, 0.25 µm) with DB-1 or DB-5 phases (J&W Scientific, USA) at a carrier gas flow-rate of 1 ml/min. The column was charged with $1-2 \mu l$ of the liquid, and the analysis was carried out with temperature programming from 50 to 250°C at a rate of 3°C/min. The chromatograms were recorded with a computer system data processing.

The values of retention times were used to calculate linear I values from Eq. (1):

$$I = \frac{(t_{\rm x} - t_{\rm n})}{t_{n+1} - t_n} \cdot 100 + 100n \tag{1}$$

Partition coefficients were calculated from the ratio of peak areas of the components by using Eq. (2):

$$K_{\rm p} = \frac{S_{\rm h}^{\rm x}}{S_{\rm a}^{\rm x}} \cdot \frac{S_{\rm a}^{\rm t}}{S_{\rm h}^{\rm t}} \tag{2}$$

where S_h^x and S_a^x are the peak areas of the component x being determined in hexane and acetonitrile and S_h^t and S_a^t are the peak areas of toluene on the chromatograms of the hexane and the acetonitrile phases, respectively.

To control the correctness of component identification, essential oils were also subjected to GC–MS analysis on an LKB-2091 instrument (Stockholm, Sweden). Separation was carried out on a capillary column with DB-5 stationary phase. Separation conditions were the same as in GC analysis with FID. The temperature of the mass spectrometer separator was 200°C and that of the ion source was 250°C. Ionisation was carried out by electrons with energy of 70 eV. The cathode current was 25 µA and the accelerating voltage was 3.5 kV.

The prediction of partition coefficients based on physicochemical properties and the measured retention indices was made by the least-squares method.

3. Results and discussion

The chromatograms in Fig. 1. illustrate the partition of essential oil components between hexane and acetonitrile phase. Table 1 gives the results of Idetermination on a DB-5 column [9] and those of partition coefficients for 70 substances, 63 of which belong to terpene compounds.

Table 2 gives the results of statistical processing of partition coefficients for different groups of structurally related compounds. The values for DB-5 and DB-1 stationary phases virtually coincide, al-



Fig. 1. Partition of the lavender essential oil components between hexane (a) and acetonitrile (b) phase. Identified components: 1 = tricyclene, $2 = \alpha$ -pinene, 3 = camphene, $4 = \beta$ -pinene, 5 = myrcene, $6 = \alpha$ -terpinene, 7 = 1, 8-cineol, 8 = limonene, $9 = \beta$ -phellandrene, $10 = \beta$ -(*Z*)-ocimene, $11 = \beta$ -(*E*)-ocimene, 12 = fenchone, 13 = linalool, 14 = camphor, 15 = borneol, 16 = isopinocampheol(?), 17 = 4-terpineol, $18 = \alpha$ -terpineol, 20 = geraniol, 21 = linalool acetate, 22 = sabinyl acetate, 23 = neryl acetate, 24 = geranyl acetate, $25 = \beta$ -caryophyllene, $26 = \beta$ -(*E*)-farnesene, $27 = \gamma$ -bisabolene(?), 28 = vetivenene(?), 29 = cadina-1, 4-diene, 30 = longiborneol (juniperol), 31 = unknown sesquiterpenoid.

256

250		
Table	1	

Partition coefficient (K_p) of some components of essential oils

Compound	I _{cal}	$I_{\rm lit}$ [9]	$K_{ m p}\pm\sigma$	n	Essential oil ^a
(I) Monoterpene hydrocarbons					
(1) Cyclofenchene	896	_	8.87	2	1
(2) Tricyclene	920±1	926	7.99 ± 0.10	3	1, 3, 5–9
(3) α-Pinen	933±2	939	7.02 ± 0.20	10	1-10
(4) Camphene	946±2	953	6.16±0.36	10	1-5, 7, 9
(5) Sabinene	971±3	976	6.50	3	1, 4, 5, 7, 8
(6) β-Pinene	974±3	980	7.14 ± 0.25	9	1-5, 7-9
(7) Myrcene	989±1	991	3.30 ± 0.20	9	1-5, 7-9
(8) 3-Carene	1010 ± 1	1011	6.70 ± 0.43	8	1 - 4
(9) α-Terpinene	1016±1	1018	4.31 ± 0.28	3	1-6
(10) Limonene	1028 ± 2	1031	4.76 ± 0.45	7	1–9
(11) β -Phellandrene	1031	1031	4.90	2	1, 2
(12) β -(Z)-Ocimene	1037±2	1040	2.99 ± 0.19	3	1, 2, 9
(13) β -(Z)-Ocimene	1046±2	1050	3.03 ± 0.07	3	1, 2, 9
(14) γ-Terpinene	1057±2	1062	4.69 ± 0.28	5	1–9
(15) Terpinolene	1088 ± 1	1089	4.78±0.15	7	1-4, 6-8
(II) Monoterpene alcohols					
(1) Linalool	1100 ± 1	1089	0.35 ± 0.07	6	7-11
(2) Fenchol, exo	1115	1117	0.32	2	4, 5
(3) Isopulegol ^b	1150	1156	0.25	1	5
(4) Isoborneol	1156±1	1156	0.36 ± 0.02	3	1, 2, 5
(5) Borneol	1166±1	1165	0.37 ± 0.02	6	1-3, 5, 9, 11
(6) Pinocampheol, iso ^b	1178	1178	0.50	2	5, 9
(7) α -Terpineol	1189 ± 1	1189	0.36 ± 0.06	8	1-5, 9, 11
(8) Nerol	1227	1228	0.30	2	6, 9
(III) Monoterpene aldehydes and ka	etones				
(1) Fenchone	1087	1087	0.59	2	5, 9, 11
(2) Thujone, <i>trans</i> ^o	1116	1114	0.60	2	4, 5
(3) Camphor	1143 ± 1	1143	0.46 ± 0.01	4	2, 3, 5, 9
(4) 3-Pinanone	1176±2	1173	0.58 ± 0.05	8	1, 3
(5) Neral (β -citral)	1241 ± 1	1240	0.38 ± 0.02	3	11
(6) Geranial (α -citral)	1271±1	1270	0.40 ± 0.03	3	10, 11
(IV) Monoterpene acetates					
(1) Linalool	1256±3	1257	0.83 ± 0.06	4	7–10
(2) Chrysanthenyl, <i>cis</i>	1261	1262	1.40	1	5
(3) Bornyl	1288±2	1285	1.26 ± 0.05	10	1-5
(4) Sabinyl, <i>trans</i>	1292±1	1291	1.21 ± 0.08	4	4, 5, 9
(5) Verbenyl, <i>trans</i>	1294	1292	0.98	1	5
(6) α -Terpenyl	1350 ± 1	1350	1.18 ± 0.05	5	I-4
(7) Neryl	1364±1	1365	1.40 ± 0.03	3	7–9, 11
(8) Geranyl	1384±1	1383	1.42 ± 0.02	3	7–11
(V) Sesquiterpenes					
(1) Thujopsene	1428 ± 3	1429	6.40	2	4, 5,
(2) β -Caryophylene	1427 ± 4	1418	9.04 ± 0.04	5	1, 2, 4, 9, 10
(3) α -Humulene	1451 ± 2	1454	8.79 ± 0.15	5	1, 2
(4) β -(<i>E</i>)-Farnesene	1482 ± 3	1477	7.97 ± 0.23	3	4, 9, 11
(5) γ -Muurolene	1489 ± 5	1480	12.08 ± 0.10	6	1, 3, 4, 10
(6) D-Germacrene	1489 ± 5	1480	7.11 ± 0.13	3	10
(7) γ -Cadinene	1510 ± 1	1513	6.02 ± 0.09	5	1, 2, 10
(8) γ -Bisabolene [®]	1517±1	1517	9.64 ± 0.61	4	1, 3, 4, 9
(9) Vetivenene ^v	1527 ± 2	1526	10.44 ± 1.03	3	4, 9
(10) Cadina-1,4-diene	1528 ± 2	1532	10.99 ± 2.00	6	1–4, 9

Table 1. Continued

Compound	I _{cal}	<i>I</i> _{lit} [9]	$K_{\rm p} \pm \sigma$	n	Essential oil ^a
(VI) Sesquiterpenoids					
(1) (Z)-Nerolidol	1537±1	1534	0.70 ± 0.02	3	11
(2) (E)-Nerolidol	1569±1	1564	0.79 ± 0.02	3	11
(3) Juniperol	1592±2	1592	0.94 ± 0.03	3	4, 5, 9
(4) Caryophylene alcohol	1570±3	1568	1.10 ± 0.06	3	1, 11
(5) Piconia (isolongi-folanone)	1620	1618	1.31	1	11
(6) γ -Eudesmol ^b	1632	1630	0.96	1	5
(7) α -Muurolol	1649±2	1645	1.09 ± 0.07	6	1-4, 10
(8) β-Eudesmol	1651	1649	0.89	2	3, 4
(B-Selinanol)					
(9) α-Cadinol	1654 ± 2	1653	0.52 ± 0.06	5	1-4, 10
(10) Coniferyl alcohol	1663	1664	1.02	2	1-4
(11) α -Santanol	1683	1678	0.88	1	11
(12) β-Bisabolol	1685	1683	0.88	2	10, 11
(13) (Z,E)-Farnesol	1700 ± 1	1697	0.78 ± 0.02	3	11
(14) Nerolidol acetate	1715	1714	2.76	1	11
(15) β -(Z)-Santalol	1720	1720	0.96	1	11
(16) (E,E) -Farnesol	1722 ± 1	1722	0.79 ± 0.02	4	10, 11
(17) $(6R,7R)$ -Bisabolene	1735	1733	0.79	1	11
(18) β -(<i>E</i>)-Santalol	1741	1741	1.05	1	11
(19) $(6S,7R)$ -Bisabolone	1742	1744	0.65	1	11
(20) β-Bisabolen-12-ol	1759	1760	0.94	1	11
(VII) Sesquiterpene epoxide					
(1) Cedranoxyde	1544	1540	3.23	1	11
(2) Caryophylene oxide	1584 ± 3	1581	4.30 ± 0.45	3	1, 11
(3) Andrane (α -cedrene epoxide)	1585	-	2.72	1	11
(VIII) Nonterpene components					
(1) Anisole, <i>p</i> -methyl	1018	1019	0.49	2	10
(2) <i>p</i> -Cymene	1026 ± 1	1026	2.29 ± 0.30	3	3, 8, 9
(3) Methyl benzoate	1092	1091	0.23	2	10
(4) Benzyl acetate	1161	1163	0.14	1	10
(5) Eugenol	1356	1356	0.68	2	10
(6) Hexenyl benzoate ^b	1587	1583	2.77	2	10
(7) 3-Hexenylphenyl acetate ^b	1626	1631	2.33	1	10

^a Essential oils: *Pinus sylvestris* (1); *Picea abies* (2); *Larix europeae* (3); *Juniperus communis* (4); *Thuja occidentalis* (5); lemon (6); tangerine (7); bergamot (8); lavender (9); ylang-ylang (10); authentic preparations (11). *b* Identified tentatively.

though *I* values can differ by 10-20 index units (in the case of DB-1 and Carbowax 20M this difference is on the average about 150 units for hydrocarbons and attains 400-500 units for alcohols).

These differences in the values of partition coefficients for different classes of compounds provide the possibility of carrying out group identification of components of complex mixtures, which is particularly important in performing 'prospective' analyses. The advantage of this approach as compared to traditional analysis on columns with different polarities is the absence of the problem of mutual assignment of peaks of the same compounds on different chromatograms. It is also important that in this method complex preliminary sample preparation including precise control of concentrations, solvent volumes, and temperature is not required.

The hexane–acetonitrile heterogenous system is not the only system suitable for determining K_p for analytical purposes. Similar results can be obtained by using the hexane–nitromethane system [10,11].

One of the factors preventing wide practical application of the above parameters is the absence of sufficiently detailed databases. This problem can be 258

Table 2

Average $K_{\rm p} \pm \sigma$ values for the principal groups of terpene compounds

Group	n	$K_{ m p}\pm\sigma$
(I) Monoterpene hydrocarbons including	12	5.27±1.81
acycylic	3	3.10±0.17
monocyclic	4	4.61±0.27
bicyclic	4	6.76±0.44
tricyclene	1	7.99
(II) Monoterpene alcohols	7	0.35±0.06
(III) Monoterpene aldehyde and ketone	6	0.50±0.10
(IV) Acetate of monoterpene alcohols including	8	1.08±0.26
acyclic	3	0.78 ± 0.09
cyclic	5	1.21 ± 0.15
(V) Sesquiterpenes	10	8.89±1.91
(VI) Sesquiterpene alcohols	14	0.84 ± 0.15
(VII) Sesquiterpene epoxide	4	3.77±0.16

solved by the calculation of the K_p values on the bases of the 'partitioning-structure' relationships similar to the 'retention-structure' relationships used for calculating *I* [12–17]. An example of these correlation equations can be given by the three-parameter equation

$$\log K_{\rm p} = a \log X + bY + c \tag{4}$$

Various physicochemical constants can be used as X and Y descriptors. They must be characterised by a monotonic change in the homological groups considered. Such constants as molar refraction, molar volume, dissolution heat, etc. may be applied. Table 3 lists the results of calculating the K_p of monoterpene hydrocarbons from Eq. (4) by using the ring numbers in the molecule as the descriptor Y and an empirical parameter δ_t proposed in [16] as the descriptor X. It is calculated according to Eq. (5):

$$\delta_{t_i}^2 = -0.02085 \cdot \frac{T_i^2}{V_i} + 58.93 \cdot \frac{T_i}{V_i} - 6892.14 \cdot \frac{1}{V_i} - 26.76$$
(5)

where T is the boiling temperature (K) at the atmospheric pressure and V is the molar volume.

The first part of Table 3 contains the initial data for calculating of the coefficients a, b, and c from Eq. (4). The second part (control set) includes

terpenes that have not been used in calculating the coefficients in Eq. (4). The average value of relative error is 3.5%, whereas the maximum value does not exceed 10%. However, the use of Eq. (4) is hindered by the fact that for many compounds boiling tem-

Table 3

Calculation of K_p for monoterpene hydrocarbons according to Eq. (4) (variable parameters *X* and *Y* are the total solubility δ_t and the number of rings *n* in the molecule, respectively)

Monoterpene	δ_{t}	п	K_p^{exp}	$K_{\rm p}^{ m calc}$	$ \Delta K_{\rm p} $
(1) Myrcene	7.86	0	3.30	3.27	0.07
(2) β -(Z)-Ocimene	8.29	0	2.99	3.12	0.13
(3) Limonene	8.28	1	4.76	4.60	0.16
(4) γ-Terpinene	8.43	1	4.69	4.53	0.16
(5) α -Pinene	8.06	2	7.02	6.93	0.09
(6) Camphene	8.25	2	6.16	6.79	0.63
(7) β-Pinene	8.27	2	7.14	6.78	0.36
				Average	0.23
Parameters of Eq. (5): $a = -0.84825$, $b = 0.16790$, $c = 1.2738$; $R = 0.988$					
Control set ^a					
(1) β -(<i>E</i>)-Ocimene	7.89	0	3.03	3.26	0.23
(2) α -Terpinene	8.21	1	4.21	4.63	0.42
(3) Terpinolene	8.56	1	4.69	4.75	0.06
(4) 3-Carene	8.36	2	6.72	6.70	0.02
				Average	0.18

^a K_{d}^{exp} values were not used for calculation above coefficients.

peratures at atmospheric pressure or density required for calculating δ_{t_i} are not known. In these cases *I* values can be used as the descriptor *X*. The average relative error for the same 12 monoterpenes is 6%, and the maximum error does not exceed 15% (*R* = 0.978). Table 4 gives the results of these calculations performed by using *I*. The second part of the table contains the calculated values of K_p for a series of monoterpenes that are not often encountered in the composition of essential oils.

Consequently, the method of group identification considered here could be easily implemented in

Table 4

Calculation of K_p for monoterpene hydrocarbons according to Eq. (4) (variable parameters X and Y are the retention index and the number of rings in the molecule, respectively)

Monoterpene	Ι	n	K_{p}^{exp}	$K_{\rm p}^{\rm calc}$	$ \Delta K_{ m p} $
(1) Myrcene	991	0	3.30	3.12	0.18
(2) β -(Z)-Ocimene	1040	0	2.99	3.19	0.20
(3) β -(<i>E</i>)-Ocimene	1050	0	3.03	3.20	0.17
(4) α-Terpinene	1016	1	4.21	4.55	0.34
(5) Limonene	1058	1	4.76	4.57	0.19
(6) γ -Terpinene	1062	1	4.69	4.63	0.06
(7) Terpinolene	1088	1	4.78	4.68	0.10
(8) α-Pinene	939	2	7.02	6.34	0.68
(9) Camphene	953	2	6.16	6.38	0.22
(10) β-Pinene	980	2	7.14	6.45	0.69
(11) 3-Carene	1011	2	6.70	6.54	0.16
(12) Tricyclene	926	3	7.99	9.08	1.09
				Average	0.34
Parameters of Eq. (5): $a = 0.42952$, $b = 0.15$	880, $c = -0.79261;$	R = 0.978			
Results of calculation					
(1) Achillene	894	0	-	2.99	
(2) allo-Ocimene	1124	0	-	3.29	
(3) allo-Ocimene, neo-	1139	0	_	3.31	
(4) α-Pyronene	944	1	-	4.40	
(5) β-Pyronene	999	1	_	4.51	
(6) α -Phellandrene	1011	1	-	4.54	
(7) β-Terpinene	1020	1	-	4.59	
(8) β-Phellandrene	1030	1	_	4.57	
(9) Sylvestrene	1032	1	-	4.58	
(10) 1-Methyl-3-iso-propylcyclohexane	1037	1	-	4.59	
(11) Isoterpinolene	1085	1	-	4.86	
(12) Sylvoterpinolene	1146	1	-	4.79	
(13) δ-Fenchene	864	2	-	6.11	
(14) ξ-Fenchene	899	2	_	6.22	
(15) β-Thujene	899	2	-	6.22	
(16) γ-Fenchene	903	2	-	6.23	
(17) Bornylene	908	2	-	6.24	
(18) ϵ -Fenchene	912	2	_	6.26	
(19) β-Fenchene	921	2	_	6.28	
(20) α-Thujene	929	2	-	6.30	
(21) α -Fenchene	951	2	_	6.37	
(22) 4-Carene	960	2	-	6.40	
(23) Sabinene	976	2	-	6.44	
(24) 2-Carene	994	2	-	6.52	
(25) Cyclofenchene	890	3	-	8.92	

practice, and databases for partitioning coefficients can be supplemented by using calculation methods.

Acknowledgements

This work was supported by the Polish State Committee for Scientific Research (KBN grants 6/ PO4G/088/10 and 125/PO6/95/08). We want also to express our gratitude to Dr. Karol Bal (Grodzisk Mazowiecki) for the samples of terpenoids.

References

- [1] I.G. Zenkevich, J. Ecol. Chem. 4 (1993) 257.
- [2] G. Guiochon, C.L. Guillemin, Quantitative Gas Chromatography for Laboratory Analyses and On-line Process Control. Elsevier, Amsterdam, Oxford, 1988.
- [3] V.G. Berezkin, V.D. Loshilova, A.G. Pankov, V.D.Yagodovskii, Khromato-Raspredelitelnyi Metod. Nauka, Moscow, 1976, (in Russian).

- [4] M. Beroza, M.C. Bowman, Anal. Chem. 37 (1965) 291.
- [5] M.C. Bowman, M. Beroza, Anal. Chem. 38 (1966) 1427.
- [6] M.C. Bowman, M. Beroza, Anal. Chem. 37 (1965) 291.
- [7] M.C. Bowman, M. Beroza, Anal. Chem. 38 (1966) 1427.
 [8] I.G. Zenkevich, A.V. Vasil'ev, Russ. J. Anal. Chem. (in
- Russian) 48 (1993) 473. [9] R.P. Adams, Identification of Essential Oil Components by
- [9] R.P. Adams, Identification of Essential Oil Components by GC–MS, Allurd, Carol Stram, IL, 1995.
- [10] I.G. Zenkevich, I.A. Tzibulskaya, Russ. J. Phys. Chem. 71 (1997) 341.
- [11] I.G. Zenkevich, Russ. J. Org. Chem. 32 (1996) 1460.
- [12] I.G. Zenkevich, L.M. Kuznezova, Collect. Czech. Chem. Commun. 56 (1991) 2042.
- [13] R. Kaliszan, Quantitative Structure–Chromatographic Retention Relationships, Wiley, New York, 1987.
- [14] K. Heberger, Chromatographia 29 (1990) 375.
- [15] M.A. Diez, M.D. Guillen, C.G. Blanco, J. Bermejo, J. Chromatogr. 508 (1990) 363.
- [16] Z. Hu, H. Zhang, J. Chromatogr. A 653 (1993) 275.
- [17] V. Isidorov, I. Zenkevich, T. Sacharewicz, Chem. Anal. (Warsaw) 42 (1997) 627.